ClinGen	Lab			2150 Pennsyl	g Lab of Washington vania Avenue NW on, DC 20037		Phone: 202-555-1	212	
Specimen Number		Pe	Specimen ripheral		Control Number	Account Number	Account Phone Number	Route	
Patient Last Name Patient First Name Patient S& Noah Patient Middle Name					Both patients have Patient Barcode identical genetic variant analysis results, therefore we are reporting				
Patient SS#		Patient Phon	Patient Phone Total Volume therefore we are reporting a single result for both.		r both.				
Age (Y/M/D) 12 y.o & 12 y.o	Date	of Birth Fen	nale & N	Fasting Tale	Please note: This NEVER happen ir	s would n real life!			
Patient Address					Indication: Sagawa (TH Deficiency) or SPR Deficiency Family History: No known family history Ethnicity: Western European Caucasian				
Date and Time Collect	ed I	Date Entered	Date a	nd Time Reported	Physician Name Jane Ferreiro, MD	NPI	Physician	ID	
Dystonia Comp	rehensiv	e Panel		Tests (Drdered				
General Comments Please send a copy of the final report to the Molecular Science/M1 Training office via Fax at (202) 555-1212									

Clinical test results for Dystonia Comprehensive Panel

GENE	RESULTS	EXPLANATION					
SPR (2p13.2)	Pathogenic variant detected:	This result supports the diagnosis of Sepiapterin reductase deficiency . This result should be interpreted in the context of clinical presentation and results of other laboratory tests (e.g., 5HIAA, 5HVA, BH4, Neopterin, etc.).					
	Arg150 Arg150Gly	A sequencing study with PCR validation has identified one copy of this reported pathogenic variation:					
ingreoory		Arg150Gly (SPR: g.6075A>G, c.448A>G, p.Arg150Gly) variation					
		gene. This encodes an alt	is an A to G change at nucle ernate residue at position 15 d to one with a small, neutral	0 1 5			
SPR (2p13.2)Pathogenic variants detected:Lys251 Lys251Ter		This result supports the diagnosis of Sepiapterin reductase deficiency . This result should be interpreted in the context of clinical presentation and results of other laboratory tests (e.g., 5HIAA, 5HVA, BH4, Neopterin, etc.).					
		A sequencing study with PCR validation has identified one copy of this reported pathogenic variation:					
		Lys251Ter (SPR: g.9120A>T, c.751A>T, p.Lys251Ter) variation.					
		The Lys251Ter variation is an A to T change at nucleotide position 9120 in the SPR gene. This forms a premature stop codon at amino acid position 251 resulting in an abnormally short or truncated protein.					
No genetic variants were detected in:		ANO3 (11p14.3-14.2) ATP1A3 (19q13.2) CIZ1 (9q34.11) DRD2 (11q23.2) GCH1 (14q22.2) GNAL (18p11.21)	HPCA (1p35.1) KCTD17 (22q12.3) PNKD (2q35) PRKRA (2q31.2) PRRT2 (16p11.2) SGCE (7q21.3)	SLC2A1 (1p34.2) SLC6A3 (5p15.33) TH (11p15.5) THAP1 (8p11.21) TOR1A (9q34.11) TOR1AIP1 (1q25.2) TUBB4A (19p13.3)			

DISCLAIMER:

Test results should be interpreted in context of clinical findings, family history, and other laboratory data. Misinterpretation of results may occur if the information provided is inaccurate or incomplete. Rare polymorphisms exist that could lead to false negative or positive results. If results obtained do not match the clinical findings, additional testing should be considered.

CLINICAL DESCRIPTION

Dystonia is a movement disorder characterized by sustained or intermittent muscle contractions causing abnormal, often repetitive movements and/or postures. Dystonic movements are typically patterned and twisting, and may be associated with tremor. Dystonia is often initiated or worsened by voluntary action and associated with overflow muscle activation. Dystonia can be classified clinically according to age of onset, body distribution, temporal pattern, and associated features (i.e., isolated dystonia – in which it is the only motor feature except tremor; combined dystonia – in which another movement disorder is present; or complex dystonia – in which other neurologic or systemic manifestations are present).

Conditions tested:

CONDITION(S)/PHENOTYPE(S)	ALSO KNOWN AS	GENE(S) TESTED
Dystonia		All listed below and:
		HPCA (1p35.1)
		KCTD17 (22q12.3)
		PARK2 (6q26)
		PNKD (2q35)
		SLC2A1 (1p34.2)
		TOR1AIP1 (1q25.2)
Autosomal dominant torsion		TUBB4A (19p13.3)
dystonia 4		
Dystonia 1	Dystonia 1, modifier of	TOR1A (9q34.11)
	Early-Onset Primary Dystonia (DYT1)	
Dystonia 10	EPISODIC KINESIGENIC DYSKINESIA 1,	PRRT2 (16p11.2)
	Familial Paroxysmal Kinesigenic Dyskinesia	
Dystonia 12	Rapid-Onset Dystonia-Parkinsonism	ATP1A3 (19q13.2)
Dystonia 16		PRKRA (2q31.2)
Dystonia 23		CIZ1 (9q34.11)
Dystonia 24		ANO3 (11p14.3-14.2)
Dystonia 25		GNAL (18p11.21)
Dystonia 5, Dopa-responsive type	DYSTONIA, DOPA-RESPONSIVE,	GCH1 (14q22.2)
	GTP Cyclohydrolase 1-Deficient Dopa-Responsive Dystonia	
Dystonia 6, torsion		THAP1 (8p11.21)
Infantile Parkinsonism-dystonia	DOPAMINE TRANSPORTER DEFICIENCY SYNDROME	SLC6A3 (5p15.33)
Myoclonic dystonia	DYSTONIA 11, MYOCLONIC	DRD2 (11q23.2)
		SGCE (7q21.3)
Segawa syndrome, autosomal	Tyrosine Hydroxylase Deficiency,	TH (11p15.5)
recessive	Tyrosine Hydroxylase-Deficient Dopa-Responsive Dystonia	
Sepiapterin reductase deficiency	Dopa-Responsive Dystonia Due to Sepiapterin Reductase Deficiency	SPR (2p13.2)

METHODOLOGY

Full gene sequencing and deletion/duplication analysis of targeted gene coding regions are performed using Next-Generation (NGS)/Massively Parallel Sequencing (MPS). All pathogenic variants and deletions/duplications are confirmed using orthogonal technologies.

PERFORMANCE

Our analytic validation study has demonstrated >99.9% sensitivity and specificity for tested mutations.